

INTRODUCTION

- Renal excretion of N¹-Methylnicotinamide (NMN), a metabolite of nicotinamide, is used as a biomarker of niacin status.
- NMN has been investigated as a phenotypic probe for several kidney transporters, including multidrug and toxin extrusion proteins (MATEs) and organic cation transporters (OCTs).
- Using NMN as an endogenous probe is advantageous over exogenously administered compounds.
- A simplified UPLC-MS/MS assay was developed for the quantitative determination of NMN in human serum and urine and validated using the FDA Guidance for Bioanalytical Method Validation.
- NMN concentrations were measured in samples from patients with chronic kidney disease to demonstrate application the method.

OBJECTIVE

- To develop and validate a method for the quantification of NMN in human samples.

METHODS

- Serum and urine samples (50µL) both underwent protein precipitation with a methanol-internal standard solution.
- Quality controls were prepared in stripped serum or synthetic urine.
- The separation of the samples was performed using an Acquity BEH Amide (2.1 mm x 50 mm, 1.7 µm) column with a BEH Amide 1.7 µm VanGuard Pre-Column.
- The mobile phase consisted of water with 0.1% formic acid (solvent A) and acetonitrile (solvent B).
- An isocratic elution at a flow rate of 0.4 mL/min was run on an Acquity UPLC I-class (Waters) with a total run time of 2 minutes.
- The analyte was detected in positive ion mode with selected reaction monitoring (SRM) using a heated electrospray ionization (HESI) source for ionization on a triple quadrupole mass spectrometer (Thermo Scientific).

Figure 1. The chemical structures of the analyte and internal standard.

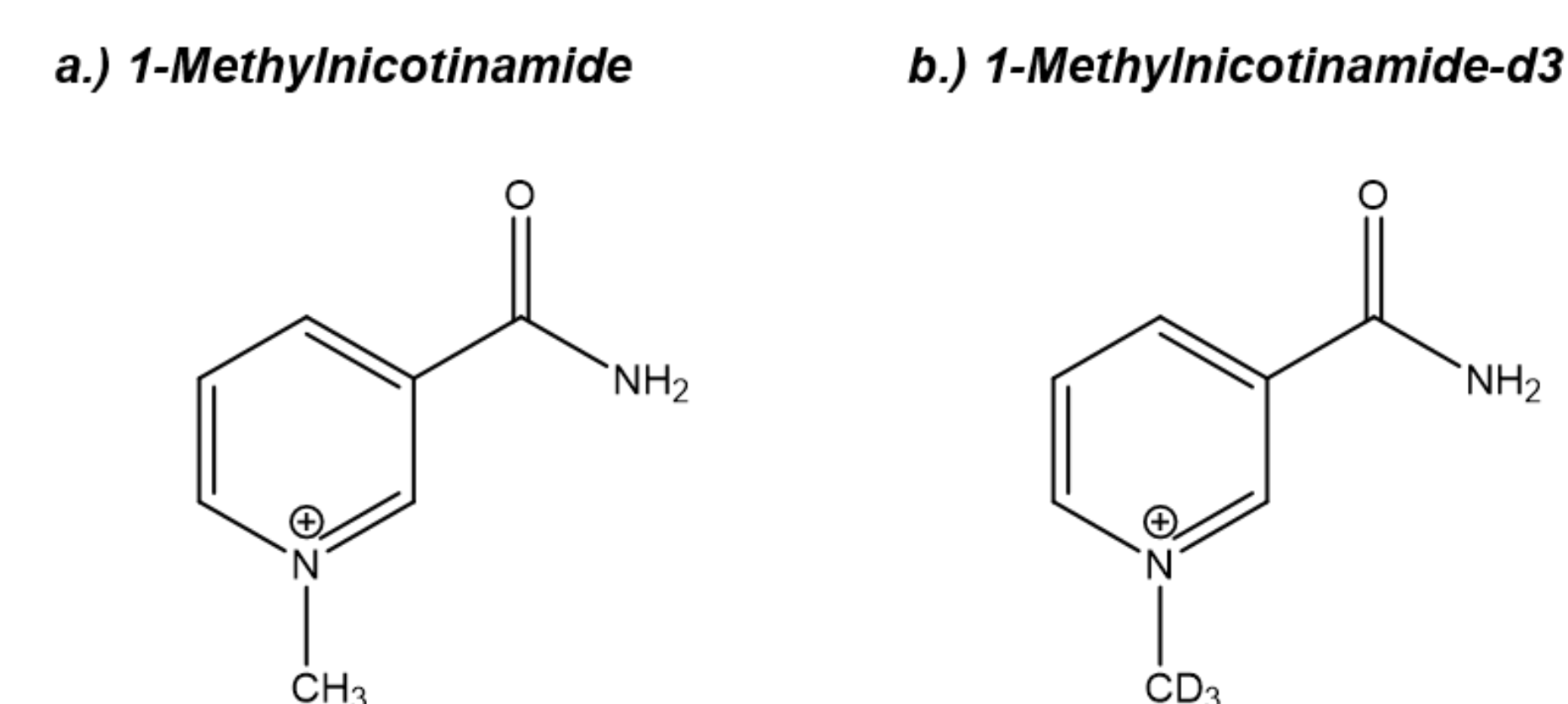


Table 1. SRM parameters for the analyte and internal standard.

Analyte	Precursor Ion (m/z)	Product Ion (m/z)	Collision Energy (V)	Tube Lens (V)
NMN	137.0	94.1	24	65
NMN-d ₃	140.0	97.2	30	86

RESULTS

Table 2. Results of the assay validation including LLOQ and linear range. The slope, intercept, and correlation coefficient are presented as mean ± standard deviation.

	Serum	Urine
LLOQ (ng/mL)	1.0 ng/mL	0.50 µg/mL
Linear range (ng/mL)	1.0-250 ng/mL	0.50-100 µg/mL
Slope (n = 3)	0.0026 ± 0.0001	0.0264 ± 0.0003
Intercept (n = 3)	0.0008 ± 0.0000	0.5638 ± 0.0080
Correlation coefficient (R ² , n = 3)	0.9976 ± 0.0016	0.9992 ± 0.0005

Table 3. Intra- and inter-day accuracy (%deviation) and precision (%CV) for LLOQ and QCs.

NMN	Level	Nominal Concentration	Intra-day ^a		Inter-day ^b	
			% Deviation	% CV	% Deviation	% CV
Serum (ng/mL)	LLOQ	1.0	1.3	10.7	0.9	9.8
	LQC	3.0	-4.1	3.8	-4.3	4.4
	MQC	75.0	-2.8	5.8	-3.3	5.8
	HQC	200	-6.7	5.4	-6.6	4.3
Urine (µg/mL)	LLOQ	0.5	1.7	3.1	-3.6	5.6
	LQC	1.5	-1.5	2.3	-4.2	4.8
	MQC	20.0	-4.9	1.9	-6.0	4.2
	HQC	80.0	-8.8	2.6	-8.3	3.1

^a Three replicates for LLOQ; 12 replicates for QCs.

^b Nine replicates for LLOQ; 24 replicates for QCs.

Table 4. Recovery and matrix effect of NMN from human serum and urine (n = 3).

NMN	QC Level	Nominal Concentration	Recovery (% mean)	Matrix Effect (% mean)
Serum (ng/mL)	LQC	3.0	100.3	99.7
	MQC	75.0	107.8	101.4
	HQC	200	98.1	97.0
Urine (µg/mL)	LQC	1.5	105.0	101.7
	MQC	20.0	97.7	92.2
	HQC	80.0	107.7	87.1

Table 5. Stability of NMN from human serum and urine (n = 3).

NMN	QC Level	Nominal Concentration	Bench Top Stability (RT, after 4 h)	Autosampler Stability (10°C, after 72 h)	Freeze/Thaw Stability (-80°C, after 3 cycles)
Serum (ng/mL)	LQC	3.0	92.1	99.7	99.0
	HQC	200	90.1	102.6	100.0
Urine (µg/mL)	LQC	1.5	104.0	105.3	103.5
	HQC	80.0	98.3	108.7	101.4

RESULTS

Figure 2. EICs for a) 1-methylnicotinamide and b) 1-methylnicotinamide-d₃ for the low, middle, and high quality control samples as well as in a human serum sample.

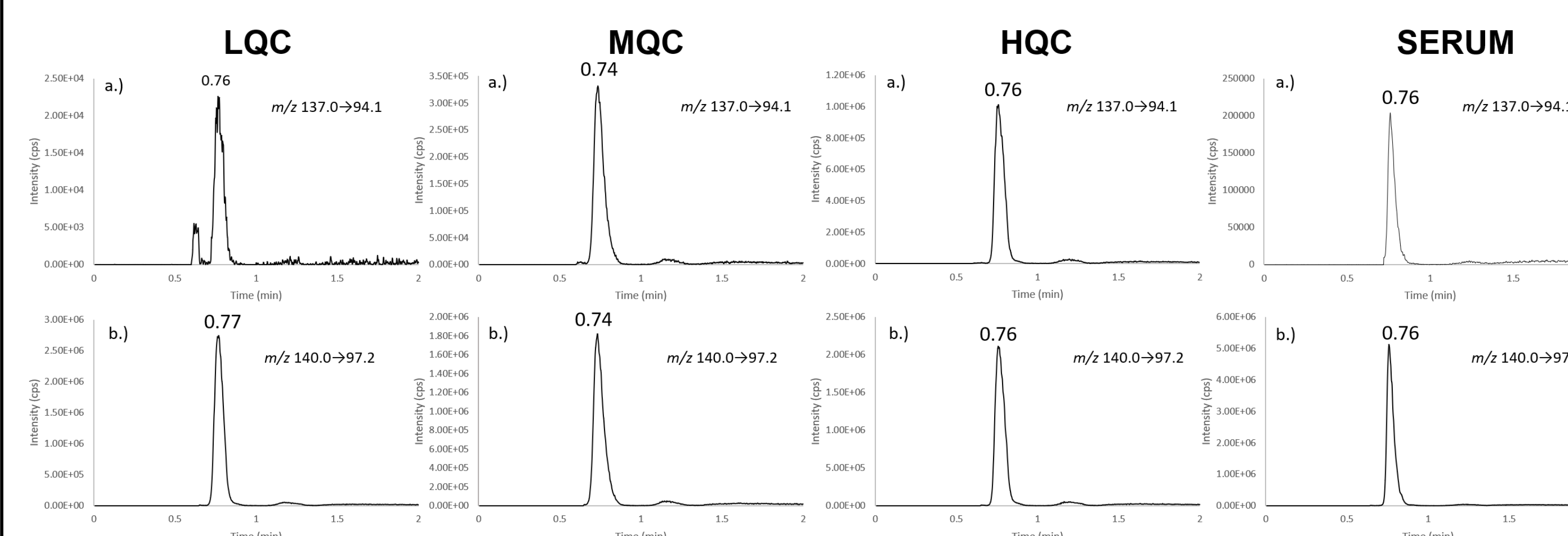
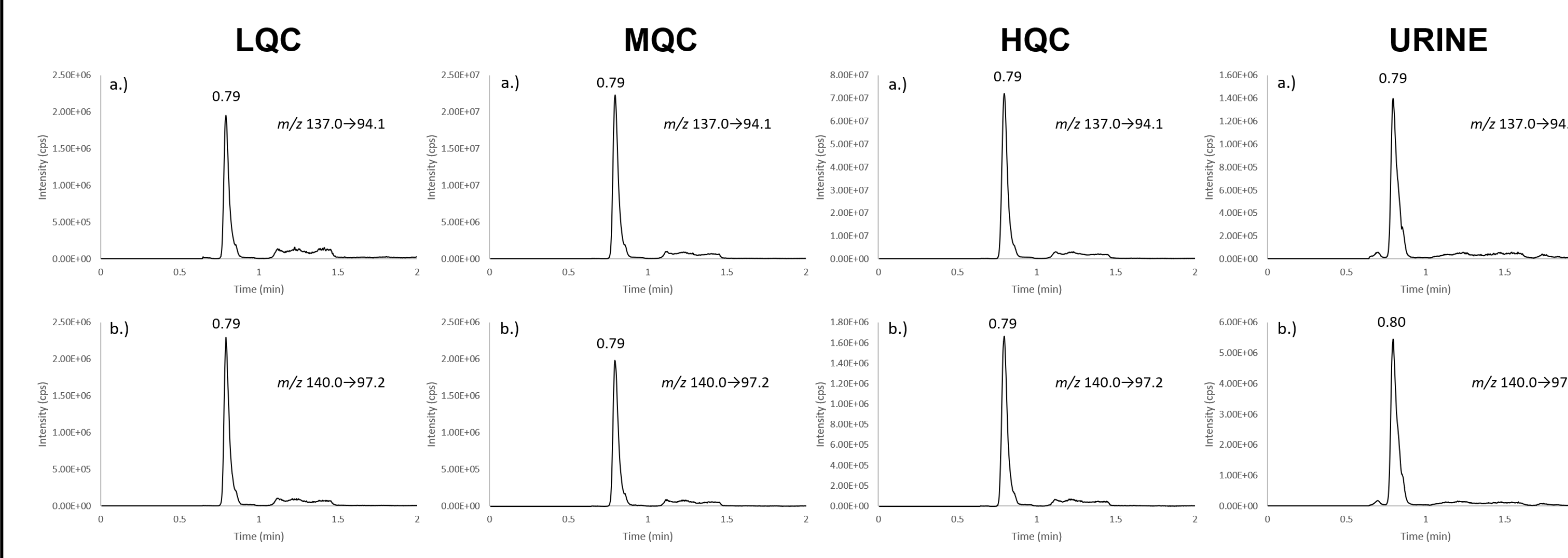


Figure 3. EICs for a) 1-methylnicotinamide and b) 1-methylnicotinamide-d₃ for the low, middle, and high quality control samples as well as in a human urine sample.



CONCLUSIONS

- A sensitive, simple, and high throughput UPLC-MS/MS assay for the quantification of NMN in human serum and urine was developed and comprehensively validated according to FDA guidelines.
- The assay has several advantages over existing assays including small sample volume requirements, minimal sample preparation, high-throughput capacity, accuracy, and precision.
- The assay is also advantageous given that NMN can be used as an endogenous probe instead of using exogenously administered compounds.
- The method was applied to measure NMN in serum and urine for a clinical study with patients with chronic kidney disease.

ACKNOWLEDGMENT

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